

Pharmacokinetics, Pharmacodynamics, and Modulation of Hepatic Glucose Production With Insulin Glargine U300 and Glargine U100 at Steady State With Individualized Clinical Doses in Type 1 Diabetes

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OBJECTIVE

This study characterized the pharmacokinetics (PK), pharmacodynamics (PD), and endogenous (hepatic) glucose production (EGP) of clinical doses of glargine U300 (Gla-300) and glargine U100 (Gla-100) under steady-state (SS) conditions in type 1 diabetes mellitus (T1DM).

RESEARCH DESIGN AND METHODS

T1DM subjects (N = 18, age 40 \pm 12 years, T1DM duration 26 \pm 12 years, BMI 23.4 \pm 2 kg/m², A1C 7.19 \pm 0.52% [55 \pm 5.7 mmol \cdot mol -1^{-1}]) were studied after 3 months of Gla-300 or Gla-100 (evening dosing) titrated to fasting euglycemia (random, crossover) with the euglycemic clamp using individualized doses (Gla-300 0.35 \pm 0.08, Gla-100 0.28 \pm 0.07 units \cdot kg⁻¹).

RESULTS

Plasma free insulin concentrations (free immunoreactive insulin area under the curve) were equivalent over 24 h with Gla-300 versus Gla-100 (point estimate 1.11 [90% Cl 1.03; 1.20]) but were reduced in the first 6 h (0.91 [90% Cl 0.86; 0.97]) and higher in the last 12 h postdosing (1.38 [90% Cl 1.21; 1.56]). Gla-300 and Gla-100 both maintained 24 h euglycemia (0.99 [90% Cl 0.98; 1.0]). The glucose infusion rate was equivalent over 24 h (1.03 [90% Cl 0.88; 1.21]) but was lower in first (0.77 [90% Cl 0.62; 0.95]) and higher (1.53 [90% Cl 1.23; 1.92]) in the second 12 h with Gla-300 versus Gla-100. EGP was less suppressed during 0–6 h but more during 18–24 h with Gla-300. PK and PD within-day variability (fluctuation) was 50% and 17% lower with Gla-300.

CONCLUSIONS

Individualized, clinical doses of Gla-300 and Gla-100 resulted in a similar euglycemic potential under SS conditions. However, Gla-300 exhibited a more stable profile, with lower variability and more physiological modulation of EGP compared with Gla-100.

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Insulin glargine 300 units \cdot mL⁻¹ (Gla-300) is a three-times concentrated glargine 100 units \cdot mL⁻¹ (Gla-100); that is, the same units are contained in one-third of the volume (1). Because of its smaller depot and surface area after subcutaneous injection, Gla-300 results into slower and more prolonged absorption, with pharmacokinetics (PK) and pharmacodynamics (PD) different from those of Gla-100 when given at the same fixed dose (0.4 and 0.6 units \cdot kg⁻¹) in subjects with type 1 diabetes mellitus (T1DM) (1). At the same fixed doses, Gla-300 mimics better than Gla-100 the physiology of basal insulin, which in the fasting state has low concentration and is flat, with no definite peak (2). When Gla-300 is compared with Gla-100 in T1DM and type 2 diabetes mellitus in the clinical EDITION program studies, Gla-300 is noninferior in glycemic control and results in a lower risk of hypoglycemia, primarily nocturnal (3-8), as well as in lower day-to-day glucose variability (9).

The slower subcutaneous absorption exposes Gla-300 to greater degradation by proteolytic enzymes (1). In fact, identical doses (0.4 units \cdot kg⁻¹ s.c.) of Gla-300 and Gla-100 resulted in lower 24-h plasma insulin exposure (by 17%) and correspondingly lower compensatory exogenous glucose infusion with Gla-300, although with a different distribution profile in T1DM (1). The PK/PD data obtained with the same, fixed insulin doses of Gla-300 and Gla-100 (1) are not necessarily applicable to the clinical situation of T1DM where subjects use individual insulin doses that are generally higher with Gla-300 to compensate for its lower bioavailability when given subcutaneously (6). Individualizing the dose of basal insulin is not only a critical way to ensure that the clamp meets its objective to produce euglycemia but also a meaningful approach to produce PK/PD data of a given basal insulin representative of the needs of the individual subject studied.

The aim of the current study was to characterize PK, PD, and the modulation of endogenous (hepatic) glucose production (EGP) at steady state (SS) with Gla-300 versus Gla-100 using the euglycemic clamp after subcutaneous injection of the individual insulin doses needed in subjects with T1DM to maintain fasting euglycemia.

RESEARCH DESIGN AND METHODS

Subjects

Subjects attending the Diabetes Clinic, Section of Endocrinology and Metabolism, Department of Medicine, University of Perugia Medical School, were enrolled and studied after giving informed written consent, provided they were diagnosed with T1DM, were aged ≥ 18 but ≤ 65 years, and with disease duration ≥ 5 years, A1C $\geq 6.5\%$ (48 mmol \cdot mol⁻¹) to $\leq 8.5\%$ (69 mmol/mol), and BMI > 20to ≤ 27 kg/m². Eighteen subjects were studied (Supplementary Table 1). They were all on a basal-bolus insulin regimen and free of detectable micro- and macrovascular complications.

Study Design

This was an investigator-initiated, randomized, single-blind (clamp investigator), two-treatment, two-period, twosequence crossover study using the euglycemic glucose clamp technique (Supplementary Fig. 1). After 2 weeks of run-in, during which subjects continued the basal-bolus regimen they were on, a computer-generated sequence was used to randomize subjects to treatment with basal insulin Gla-100 (Lantus SoloStar pen; Lantus; Sanofi, Paris, France) or Gla-300 (Toujeo, Toujeo SoloStar pen; Sanofi), both evening dosing, for 3 months. During this period, the subjects titrated basal and prandial insulin continuing the basal-bolus regimen (see below). At the end of the 3 months, subjects underwent the first euglycemic glucose clamp after (evening) a subcutaneous injection of the individual dose of Gla-100 or Gla-300 they were on during the previous 2 weeks. Subjects then underwent a 2-month washout, resuming the basal insulin treatment they were on before randomization. After the subjects were crossed over to the other basal insulin for 3 more months, during which the doses were titrated as in the first period, they were then studied with the second euglycemic glucose clamp following the same procedure as with the first clamp. The study protocol was approved by the Ethical Study Committee of Umbria Region (CEAS) and registered in EudraCT (2015-002135-17). EudraCT, European Union Drug Regulating Authorities Clinical Trials, is the European Clinical Trials database of all clinical trials of investigational

medicinal products with at least one site in the EU.

Basal Insulin Titration

Basal insulin titration was an open-label process once-twice per week based on frequent face-to-face or telephone (including e-mail, smartphone applications) contacts during both treatment periods. At baseline, all subjects were familiar with frequent self-monitoring of blood glucose (SMBG) (six to eight plasma glucose [PG] measurements daily before and 2 h after each meal, midafternoon, and bedtime, and at least twice weekly at 0300 h). After randomization, Gla-100 and Gla-300 (dosing at 2000 h, \pm 30 min) were both titrated aiming at fasting PG (FPG) between 90 and 110 mg \cdot dL⁻¹. Dose change decision was made no more frequently than 3-4 days based on the difference between bedtime and fasting PG on days in which postdinner PG was 100-130 $mg \cdot dL^{-1}$ (Supplementary Table 2) (10).

Clamp Procedure

The principle of the hyperinsulinemiceuglycemic glucose clamp was used in the current study (Supplementary Fig. 2) (11). The euglycemic feedback (12) and clamp procedure for the purpose of examining PK/PD of long-acting insulin analogs has previously been described in detail (13-15). The study was blind to the clamp investigators who were not aware of the basal insulin treatment the subjects were on. In brief, subjects were admitted to the Clinical Research Center of Department of Medicine, University of Perugia Medical School, between 1430 and 1500 h of the first study day, \sim 1.5 h after the end of their usual lunch preceded by a subcutaneous injection of rapid-acting insulin dose. They were put at bedrest and maintained supine and fasted until 2000 h the next day. Two superficial venous lines were started. A hand vein of one arm was cannulated retrogradely with a 21-gauge butterfly needle, and the hand was maintained in a hot box or pad (\sim 65°C) for intermittent sampling of arterialized venous blood (16) for PG and hormone measurements. A superficial vein of the contralateral arm was cannulated with an 18-gauge catheter needle for an intravenous (i.v.) infusion of insulin and/or 20% dextrose solution as needed. At 1500 h, a feedback i.v. infusion of human regular insulin (regular insulin [Eli Lilly Italia SpA], diluted to 1 unit \cdot mL⁻¹ in 100 mL saline solution containing 2 mL of the subject's blood), was initiated, whenever needed, using a syringe pump (Injectomat MC Agilia; Fresenius Kabi, Hessen, Germany) to maintain PG at 100 mg \cdot dL⁻¹ according to a modified algorithm (13).

At -180 min from time "0 min," (t_0) a primed sterile, pyrogen-free constant infusion (0.222 μ mol \cdot kg⁻¹ \cdot min⁻¹) of [6,6-²H₂]glucose (Cambridge Isotopes Laboratories, Cambridge, MA) was started and maintained throughout to determine glucose kinetics. During the clamp, a variable rate of infusion of 20% glucose with 2% [6,6-2H2]glucose was used to avoid non-SS errors (17) in the measurement of glucose turnover, as described previously (18). A peristaltic pump (Volumat MC Agilia; Fresenius Kabi) was used for infusion of glucose, whenever needed, to prevent fall of PG below 95 mg \cdot dL⁻¹ during feedback and clamp periods. At 2000 h, the dose of basal insulin, either Gla-100 or Gla-300, the individual subjects were receiving over the last 15 days, was injected subcutaneously 2 cm to the right or left of the umbilicus by a SoloStar pen (4-mm needle, skinfold technique), and subjects underwent the euglycemic clamp for 24 h.

Analytical Methods

Bedside PG was measured in triplicate every 5-15 min using the YSI 2300 STAT glucose analyzer (Yellow Springs Instruments, Yellow Springs, OH). Plasma C-peptide was measured by radioimmunoassay (Linco Research, St. Charles, MO). The plasma free insulin concentration (free immunoreactive insulin [FIRI]) was measured by a commercial radioimmunoassay kit specific for human insulin (DRG Instruments GmbH, Marburg, Germany) with a range of detection of 0–200 μ U \cdot mL⁻¹ after polyethylene glycol extraction of antibodies from the plasma (19). A1C was determined by high-performance liquid chromatography using an HI-Auto A1c HA-8121 apparatus (DIC, Daiichi, Kogaku Co., Ltd., Kyoto, Japan) that was Diabetes Control and Complications Trial (DCCT) aligned (upper limit in subjects without diabetes <6.1%). To determine glucose kinetics, arterialized venous blood samples were taken every 30 min for the first 6 h and then every 60-120 min throughout the studies. Glucose enrichment was determined on its penta-acetate (penta-*O*-acetyl- β -D-glucopyranose) derivative by gas chromatography-mass spectrometry (GC HP 6890 II, MS HP 5973/A; Hewlett-Packard Co., Palo Alto, CA) in electron impact ionization mode monitoring the ions 200 and 202 for the unlabeled and D-[6,6-²H₂]glucose, respectively (20).

End Points

The primary study end point was the glucose infusion rate area under the curve (GIR-AUC) versus area under the time curve 0-24 h (GIR-AUC_{0-24 h}). Secondary PD end points included GIR-AUC(0-12 h), GIR-AUC(12-24 h), GIR-AUC(0-6 h), GIR-AUC(6-12 h), GIR-AUC(12-18 h), GIR-AUC(18-24 h), maximum GIR (GIR_{max}), time to 50% of $GIR-AUC_{(0-24 h)}$, and GIR fluctuations. Other secondary PK end points included FIRI-AUC(0-24 h) and the various time intervals for FIRI: FIRI-AUC(0-12 h), FIRI-AUC(12-24 h), FIRI-AUC(0-6 h), FIRI-AUC(6-12 h), FIRI-AUC(12-18 h), FIRI-AUC(18-24 h), maximum FIRI concentration (C_{max}), time to 50% of FIRI-AUC(0-24 h), swing [(FIRI-Cmax -FIRI-C_{min})/FIRI-C_{min}], fluctuation, and insulin excursions. Other secondary end points were glucose fluxes (EGP, glucose utilization [GU]).

Calculations and Statistical Analyses

The linear trapezoidal rule was used to calculate the AUC for FIRI and GIR (untrasformed data). GIR data were smoothed by taking a three-point moving average to provide data for calculation of GIRmax. Cmax was read directly from the plasma concentration-time data for each subject. To express within-day insulin concentration variability, the following indices were calculated: swing ([C_{max} - C_{min}]/ C_{min}), fluctuation over 24 h (F24) in insulin concentration (quantified as cumulated AUCs of an individual's FIRI profile above and below the average FIRI over 24 h), relative degree of fluctuation (100% \times [F24/C_{avg}]) and insulin excursion ($C_{max} - C_{min}$). Within-day GIR fluctuation around the average value was calculated as for FIRI, as previously reported (1). All calculations were made on untransformed data.

Glucose fluxes were calculated based on a non-SS assumption, and the total R_a and R_d were calculated by using a modified form of the Steele equation to account for the addition of stable labeled tracer to the exogenous glucose infusate (17). EGP (primarily hepatic) was obtained as the difference between R_a and the exogenous GIR during the clamp. GU was obtained by adding the GIR to the EGP (21). When the EGP yielded a negative number, EGP was assumed to be zero, and the corresponding GIR was assumed as GU.

Point estimates of treatment ratios (Gla-300-to-Gla-100), with 90% Cls, were calculated using ANOVA, which allowed for variation due to sequence, subjects nested within sequence, period, and treatment based on log-transformed data and retransformations. Time to 50% of GIR-AUC_(0-24 h) and time to 50% of FIRI-AUC_(0-24 h) were analyzed nonparametrically using the Wilcoxon rank sum test and Hodges-Lehmann estimates of the treatment differences computed with 90% Cls. Equivalence was to be concluded if the 90% CIs for the PK/PD AUC and C_{max} were completely contained within the interval 0.80-1.25. A total sample size of 18, using two onesided tests on data from a two-period crossover design, was calculated to provide at least 80% power at a 5% significance level, assuming a true ratio of the means of 1.0 and the coefficient of variation of 0.22, to demonstrate that the 90% CI of the ratios of key PK or PD parameters between treatments would be contained within 0.80-1.25. Data are expressed as arithmetic means (SD), geometric means (95% CI) in tables, and as means and SE in figures. Statistical analysis was usually performed using NCSS 12/PASS 11 software (NCSS, LLC, Kaysville, UT).

RESULTS

Glycemic Control, Body Weight, and Insulin Doses Before Studies

Mean daily blood glucose over the week before studies (SMBG) with Gla-300 and Gla-100 was similar (149 \pm 17 and 152 \pm 19 mg· dL⁻¹, respectively). No hypoglycemia (PG \leq 70 mg \cdot dL⁻¹) was reported by SMBG during the 3 days before the studies. A1C similarly decreased from baseline to the end of the 3-month period of treatment with both Gla-300 and Gla-100 (Supplementary Table 1). Body weight did not change significantly from baseline to the end of treatment with Gla-300 and Gla-100 (Supplementary Table 1). The dose of basal insulin at the end of the 3-month period was higher with Gla-300 versus Gla-100, with a difference of 0.071 units \cdot kg⁻¹ (90% CI 0.059; 0.083), whereas the dose of prandial insulin did not change, although the percentage of prandial insulin was lower with Gla-300 versus Gla-100 (Supplementary Table 1).

Intravenous Insulin and Glucose Infusion and PG Concentration Before Clamp Studies

On the first study day, PG at 1500 h (\sim 1.5 h after end of lunch and 5 h before subcutaneous injection of basal insulin

and clamp initiation at 2000 h (t_0) was similar with Gla-300 and Gla-100, and decreased slowly to euglycemia by 80 min before t_0 , with no difference between the two insulins (Fig. 1). When appropriate, a low dose of i.v. insulin infusion to reach euglycemia was started between 5 and 4.5 h before t_0 and continued until 1.5 h before t_0 and continued until 1.5 h before t_0 when it was virtually zero. The total amount of insulin infused (insulin infusion rate) before t_0 was lower with Gla-300 than with Gla-100 (insulin infusion rate-AUC_(-5 to 0 h),

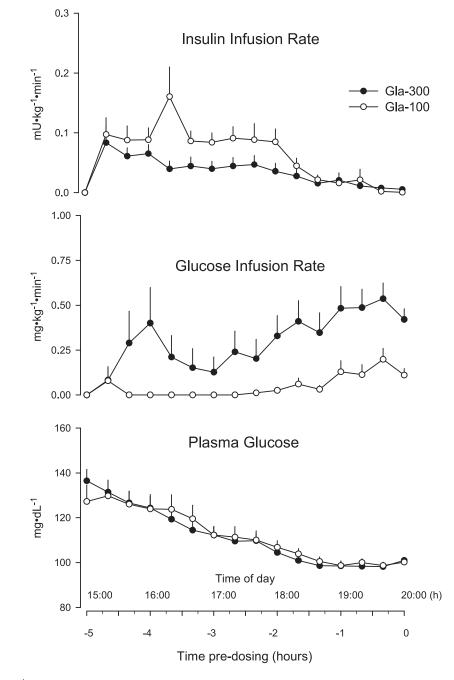


Figure 1—Rates of i.v. insulin and glucose infusions and PG concentration in the 5 h (1500–2000 h) before initiation of the clamp studies at t_0 (2000 h).

11.5 \pm 7.3 vs. 19.5 \pm 9.9 μ U · kg⁻¹; 0.57 [95% CI 0.39; 0.81]). When needed, a glucose infusion was started 4.5 h before t_0 . GIR increased more with Gla-300 than with Gla-100 until t_0 (GIR-AUC _(-5 to 0 h), 90.1 \pm 99.3 vs. 14.5 \pm 17.9 mg · kg⁻¹; 7.9 [90% CI 4.17; 15.12]).

Plasma Insulin and Glucose Concentrations and GIR After Subcutaneous Injection of Gla-300 and Gla-100

At t_0 , plasma insulin concentration was higher with Gla-300 versus Gla-100 (10.2 \pm 1.0 $\mu\text{U}\cdot\text{mL}^{-1}$ and 8.2 \pm 1.4 μU · mL⁻¹; 1.38 [90% CI 1.15; 1.66]). During the first 6 h after the subcutaneous insulin injection, insulin concentration increased less with Gla-300 than after Gla-100 (FIRI-AUC_(0-6 h), 0.91 [90% CI 0.86; 0.97]) (Table 1). Thereafter, plasma insulin plateaued similarly with Gla-300 and Gla-100 until 12 h, after which time FIRI-AUC(12-24) was 38% greater with Gla-300 until the end of the study (Table 1 and Fig. 2). The maximal serum insulin concentration was lower with Gla-300 versus Gla-100 (0.87 [90% CI 0.81; 0.92]) (Table 1). All indices of the within-day variability of plasma insulin concentration (swing, both absolute and relative fluctuation, and insulin excursion) indicated that this was lower by \sim 50% with Gla-300 compared with Gla-100 (Table 1).

GIR at t_0 was greater with Gla-300 versus Gla-100 (0.42 \pm 0.25 vs. 0.11 \pm $0.15 \,\mathrm{mg} \cdot \mathrm{kg}^{-1} \cdot \mathrm{min}^{-1}$ (median difference, 0.3 [90% CI 0.17; 0.45]). The overall glucose infused to maintain euglycemia was no different between Gla-300 and Gla-100 over the full 24-h study period (GIR-AUC_(0-24 h), 1.03 [90% CI 0.88; 1.21]) (Table 1 and Fig. 2). However, after the subcutaneous injection, GIR increased less during the first 12 h with Gla-300 compared with Gla-100 (GIR-AUC_(0-12 h), 0.77 [90% CI 0.62; 0.95]) (Table 1), with the greatest difference observed in the first 6 h (GIR-AUC(0-6 h), 0.71 [90% CI 0.56; 0.91]) (Table 1). Thereafter, GIR was similar up to 18 h, when the trend reversed with higher glucodynamic effect for Gla-300 compared with Gla-100 (GIR-AUC(18-24 h), 1.91 [90% CI 1.37; 2.68]) (Table 1, Fig. 2, and Supplementary Fig. 3). The maximal GIR was lower by 15% with Gla-300 versus Gla-100 (Table 1 and Fig. 2). Similarly, absolute and relative

| | 0.35 units \cdot kg ⁻¹ | 0.28 units \cdot kg ⁻¹ | Gla-300-to-Gla-100 ratio |
|---|-------------------------------------|-------------------------------------|--------------------------------------|
| | Gla-300 | Gla-100 | Point estimate ^a (90% CI) |
| PK parameters | | | |
| FIRI-AUC _{0-24 h} (μ U · h ⁻¹ · mL ⁻¹)*^ | 257 (208; 317) | 231 (182; 294) | 1.11 (1.03; 1.20) |
| FIRI-AUC _{0-12 h} (μ U · h ⁻¹ · mL ⁻¹)* | 139 (115; 168) | 145 (116; 181) | 0.96 (0.89; 1.04) |
| FIRI-AUC _{12-24 h} (μ U · h ⁻¹ · mL ⁻¹)* | 116 (91; 148) | 84 (63; 113) | 1.38 (1.21; 1.56) |
| FIRI-AUC _{0-6 h} (μ U · h ⁻¹ · mL ⁻¹)* | 71 (58; 86) | 78 (63; 96) | 0.91 (0.86; 0.97) |
| FIRI-AUC _{6-12 h} (μ U · h ⁻¹ · mL ⁻¹)* | 68 (57; 82) | 67 (52; 86) | 1.03 (0.92; 1.14) |
| FIRI-AUC _{12–18 h} (μ U · h ⁻¹ · mL ⁻¹) | 59 (47; 75) | 48 (36; 63) | 1.24 (1.20; 1.38) |
| FIRI-AUC _{18–24 h} (μ U · h ⁻¹ · mL ⁻¹)* | 56 (42; 74) | 36 (25; 51) | 1.58 (1.31; 1.88) |
| FIRI-C _{max} (μ U · mL ⁻¹)* | 14.5 (12.1; 14.4) | 16.7 (13.5; 20.8) | 0.87 (0.81; 0.92) |
| T _{50%} –FIRI-AUC _{0–24} (h)§ | 10 (7; 12) | 8.5 (7; 12) | 0.5 (0; 1.0) |
| Swing ^b | 1.0 (0.7; 1.4) | 3.1 82.3; 4.2) | 0.33 (0.26; 0.43) |
| Fluctuation $(\mu U \cdot mL^{-1})^{c}$ | 0.65 (0.5; 0.8) | 1.20 (1.1; 1.4) | 0.54 (0.45; 0.64) |
| Fluctuation (%) ^d | 6.0 (4.1; 8.6) | 12.0 (8.5; 16) | 0.51 (0.45; 0.64) |
| Δ INS (μ U · mL ⁻¹) ^e | 7.1 (6; 8.3) | 12.3 (10; 15) | 0.57 (0.49; 0.67) |
| PD parameters | | | |
| $GIR-AUC_{0-24 h} (mg \cdot kg^{-1})^*$ | 981 (803; 1,198) | 950 (758; 1,190) | 1.03 (0.88; 1.21) |
| $GIR-AUC_{0-12 h} (mg \cdot kg^{-1})^*$ | 464 (345; 613) | 604 (476; 766) | 0.77 (0.62; 0.95) |
| $GIR-AUC_{12-24 h} (mg \cdot kg^{-1})^*$ | 482 (394; 590) | 314 (228; 433) | 1.53 (1.23; 1.92) |
| $GIR-AUC_{0-6 h} (mg \cdot kg^{-1})*$ | 251 (172; 366) | 353 (292; 426) | 0.71 (0.56; 0.91) |
| $GIR-AUC_{6-12 h} (mg \cdot kg^{-1})^*$ | 165 (98; 277) | 229 (153; 344) | 0.72 (0.44; 1.18) |
| $GIR-AUC_{12-18 h}$ (mg \cdot kg ⁻¹) | 137 (72; 261) | 149 (88; 253) | 0.92 (0.59; 1.44) |
| $GIR-AUC_{18-24 h} (mg \cdot kg^{-1})^*$ | 280 (230; 341) | 144 (101; 205) | 1.91 (1.37; 2.68) |
| $GIR-C_{max}$ (mg · kg ⁻¹ · min ⁻¹)* | 1.5 (1.3; 1.7) | 1.7 (1.5; 2.0) | 0.85 (0.79; 0.91) |
| T _{50%} –GIR-AUC _{0–24} (h)§ | 11.7 (6.3; 18) | 8.3 (4; 13.7) | 3.35 (1.85; 5.15) |
| Fluctuation (mg \cdot kg ⁻¹ \cdot min ⁻¹) ^c | 0.61 (0.5; 0.8) | 0.73 (0.6; 0.9) | 0.83 (0.74; 0.93) |
| Fluctuation (%) ^d | 82 (65; 106) | 102 (84; 124) | 0.81 (0.70; 0.94) |

Table 1—PK and PD variables after subcutaneous injection of individual doses of insulin Gla-300 and insulin Gla-100 in subjects with T1DM in SS

Within-day GIR fluctuation around the average value was calculated as for FIRI. ^aPoint estimates of treatment ratios with 90% CIs were calculated using a linear mixed-effects model based on log-transformed data and retransformations. *Data are geometric mean (95% CI). §Data are median (min; max) (N = 18). ^FIRI (serum insulin was measured after extraction of antibodies with 30% polyethylene glycol). §Point estimates for treatment effects are based on the Hodges-Lehmann estimate of the median difference with the associated 90% CI. ^bSwing: [(FIRI C_{max} – FIRI C_{min})/FIRI C_{min}]. ^cFluctuation (F24): [(FIRI C_{max} – FIRI C_{min})/C_{avg}]. ^dFluctuation (%): [(100% × F24)/FIRI C_{avg}]. ^e Δ INS: (FIRI C_{max} – FIRI C_{min}).

fluctuations in GIR around the average were both lower, by 17% and 19%, respectively, with Gla-300 compared with Gla-100 (Table 1).

The PG concentration was no different during the 24-h clamp with Gla-300 (100.5 \pm 1.2 mg \cdot dL⁻¹) and Gla-100 (101.4 \pm 1.8 mg \cdot dL⁻¹; 0.99 [90% CI 0.98; 1.0]).

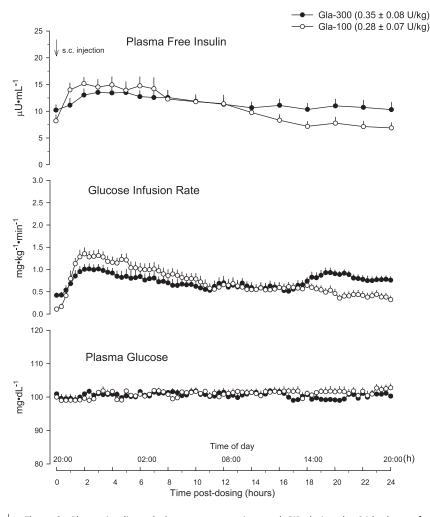
Glucose Fluxes (EGP and GU)

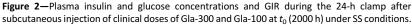
Data were available from 14 of the 18 subjects studied (the glucose enrichment was insufficient in 1 subject, the samples were lost in one study in another subject, and the isotope was not infused in 2 subjects). The anthropometric characteristics of the 4 subjects for whom data were missing were not different from those of the rest of the study group. Supplementary Fig. 4 shows the tracerto-tracee ratio (TTR) in study. The changes of TTR over the 24 h (as well as in the first and second 12-h periods) with Gla-300 and Gla-100 were no different (two-way repeated-measures ANOVA

[treatment × time, $F_{15,195}$ = 1.79; P = 0.177, Greenhouse-Geisser correction; power 0.92, with α = 0.05]) At t_0 , EGP was lower with Gla-300 (1.85 \pm 0.82 vs. 2.07 \pm 0.52 mg \cdot kg⁻¹ \cdot min⁻¹, respectively; P < 0.048). After the subcutaneous injection of basal insulin, EGP decreased both with Gla-300 and Gla-100 as expected. Suppression of EGP was less with Gla-300 compared with Gla-100 in the first 6-h interval (EGP-AUC_{IO-6 hl}, ratio 1.41 [90% CI 1.01; 1.95]), but greater in the 18-24-h interval (EGP-AUC_[18-24 h], ratio 0.57 [90% CI 0.36; 0.92]) (Fig. 3 and Supplementary Fig. 3). However, over the 24-h period, EGP with Gla-300 was similar to that of Gla-100 (ratio 0.98 [90% CI 0.80; 1.21]). GU did not increase after subcutaneous Gla-300 or Gla-100 and was no different with between the two treatments (Fig. 3).

CONCLUSIONS

The current study was undertaken to establish the PK/PD as well as the modulation of EGP using clinical, individual doses of Gla-300 compared with Gla-100 under experimental conditions mimicking as close as possible the clinical situation where subjects with T1DM are usually on different doses of the two insulins (6,7). Such a study design is unprecedented, with one exception (22). Under these conditions, the doses of insulin differ not only between individuals but also in the same individual switching from Gla-100 to Gla-300 and vice versa. In the current study, after a 3-month titration/optimization period, the same glycemic control (A1C) was achieved with 0.07 units \cdot kg⁻¹ higher dose of Gla-300 compared with Gla-100, a result consistent with Comparison of a New Formulation of Insulin Glargine With Lantus in Patients With Type 1 Diabetes Mellitus (EDITION 4) (evening dosing group) (6). When these different insulin doses of Gla-300 and Gla-100 were tested in the current study in the same individuals, the two insulins exhibited similar total exposure and required a similar amount of infused glucose over





24 h, but there were differences in the profile. Indeed, the distribution of exposure and effect was more balanced between the first and the second 12-h period of the 24-h study with Gla-300, and thus, the within-day PK/PD variability of Gla-300 was lower than with Gla-100. In addition, Gla-300 appeared to be more consistent in modulating EGP both in the first and the second 12-h period of the 24-h study than Gla-100.

Regarding PK, the current study, with a higher dose of Gla-300 versus Gla-100, originally indicates that the mean plasma insulin concentration over 24 h did not differ (Table 1). This finding is different from that reported with same fixed Gla-300 and Gla-100 dose of 0.4 units \cdot kg -1^{-1} , where plasma insulin over 24 h was lower with the former versus the latter (1). In the current study, there were important differences in plasma insulin profiles between the first and the second 12 h of the 24-h study. Plasma

insulin increased less with Gla-300 than with Gla-100 over the first 6 h but remained steady and was consistently greater with Gla-300 than with Gla-100 in the 12- to 24-h interval postdosing (Table 1 and Fig. 2). Overall, the 24-h PK profile of plasma insulin with Gla-300 appeared flatter and more constant over the 24-h observation period than with Gla-100 (Fig. 2). Notably, the 24-h postinjection plasma insulin concentration of Gla-300 was similar to that at predosing at t_0 , a value reflecting Gla-300 given the day before day 1 of the study, suggesting consistency (low day-to-day variability) of the same clinical insulin doses of Gla-300, whereas it was lower with Gla-100 (Fig. 2).

The total GIR-AUC over the 24 h of study with Gla-300 and Gla-100 was no different (Table 1). This also is an original finding, which is at variance with the result of less glucose-lowering effect reported with Gla-300 compared with same fixed dose of Gla-100 (1) and establishes equivalence in 24-h PD for individually titrated clinical doses of Gla-300 versus Gla-100. However, there was less variability in glucodynamics (GIR) with Gla-300. In parallel with PK, GIR increased less after the subcutaneous injection of Gla-300 compared with Gla-100 for the first 12 h of the study. Then, GIR was similar between 12 and 18 h and ultimately greater between 18 and 24 h with Gla-300 versus Gla-100.

As expected from the physiology of the insulin dose-response relationship on glucose fluxes (23), the marginal increase in the plasma insulin concentration after the subcutaneous injection of Gla-300 or Gla-100 did not stimulate peripheral GU (primarily insulin-mediated muscle uptake) but had effects especially on suppression of EGP (Fig. 3). The lower suppression of EGP in the initial part of study and its more consistent suppression in the late part, with Gla-300 versus Gla-100, indicates a more physiological modulation of EGP with the former compared with the latter basal insulin. The absolute values of EGP observed in this study should be interpreted with caution due to the unprecedented infusion of the tracer for 24 h (24). Previous observational studies in normal (25) and T1DM subjects (26) have reported an increase in TTR and underestimation by \sim 30% of EGP after longversus short-term infusion of the tracer. This has been interpreted as the result of glucose recycling (in the current study possibly from glycogen via incorporation and release of [6,6-²H₂]glucose) (26) and/ or failure to achieve complete equilibrium of the tracer with the tracee during the short infusion (25). In the present studies, the TTR increased slowly for the initial 8 h; then, interestingly, it decreased in the early morning hours and, ultimately, increased again in the last part of the study (Supplementary Fig. 4). However, the changes of TTR were superimposable with the two insulins, indicating that even if there were some tracer recycling, this was most likely similar with Gla-300 and Gla-100, and the relative differences of EGP observed with Gla-300 and Gla-100 are real. This interpretation is independently supported by the GIR changes, which, in the present experimental model of hyperinsulinemic-euglycemic clamp with low plasma insulin concentrations and

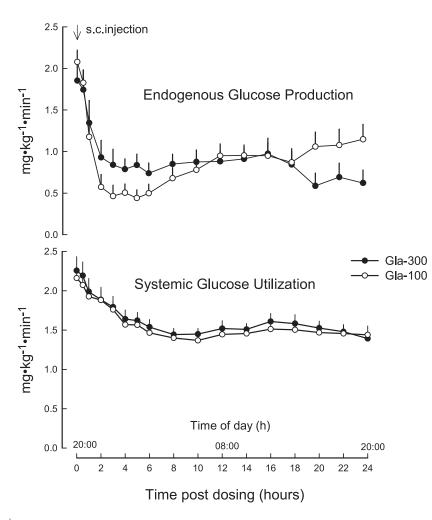


Figure 3—Rates of EGP and systemic GU during the 24 h after subcutaneous injection of clinical doses of Gla-300 and Gla-100 at t_0 (2000 h) under SS conditions.

no increase in muscle glucose uptake, reflect primarily changes in EGP (23).

The differential modulation of EGP, the primary determinant of PG concentration with Gla-300 compared with Gla-100, might translate into practical implications for subjects with T1DM. On one hand, the lower and smoother suppression of EGP in the initial part of the current study with evening dosing of Gla-300 versus Gla-100 possibly reduces the risk for nocturnal hypoglycemia at this vulnerable time of day (27) with former compared with latter basal insulin, a result reported by the EDITION studies with evening dosing (3-8). On the other hand, the more consistent suppression of EGP with Gla-300 versus Gla-100 18-24 h postdosing predicts less hyperglycemia in the afternoon with the former than with the latter. This effect may be beneficial to those T1DM subjects who present hyperglycemia in the late afternoon and are sometimes given basal insulin twice daily (28). In the EDITION JP 1, Gla-300 resulted in lower predinner PG versus Gla-100 (29).

In the current study, the within-day variability of Gla-300 was lower compared with Gla-100 both for PK and PD (Table 1). Swing and fluctuation of PK were lower with Gla-300, and fluctuation of PD was also lower with Gla-300 compared with Gla-100. Similar results have been previously reported, however, in healthy subjects (30) and with the same fixed doses of insulin (30–32).

The PK and PD data of the current study are qualitatively consistent with those of Becker et al. (1). However, the current study design makes the results more directly applicable to the real life of T1DM subjects than those of the fixed doses by Becker et al. (1). In addition, the population in the current study was more homogeneous, with all subjects at run-in on the same basal insulin (Gla-100). In contrast to previous studies, the current study included a relevant (predominant) number of female subjects who were studied in the follicular phase of their menstrual cycle on both occasions.

In the current study, great care was given to the preparation of subjects immediately before the clamp experiments to achieve similar metabolic conditions before the Gla-300 and Gla-100 study, as shown in Fig. 1. Ensuring that subjects are in reproducible metabolic conditions at t_0 of the clamp studies with a crossover study design is, of course, important for reliable interpretation of data generated.

The results of the current study were obtained with evening dosing. Additional studies are needed to examine PK/PD of Gla-300 and Gla-100 given in the morning (a more and more attractive time of injection for patients) (6,9,30) to establish whether differences in circadian rhythm (18,33–35) may affect the PK/PD observed in the current study with evening dosing.

The current study has limitations such as the number of subjects studied and the single-blind not double-blind study design. Strengths are the PK/PD data generated in a homogeneous group of subjects, including women, in fair glycemic control, all treated with the same basal insulin Gla-100 at baseline, the reproducible metabolic conditions before paired clamp studies, and the novel approach to study PK/PD of Gla-300 and Gla-100 with individually titrated clinical doses of each insulin resulting in matched glycemic control.

In conclusion, the study at SS indicates that PK/PD of both Gla-300 and Gla-100, titrated to fasting euglycemia in subjects with T1DM, have similar glucoselowering effects over 24 h. However, evening dosing of Gla-300 exhibits potentially favorable differences in PK/PD profiles and in modulating EGP that result in lower insulin activity at night, greater insulin activity in the afternoon, and lower within-day PK/PD variability. Thus, evening dosing of Gla-300 has the potential of reducing the risk of nocturnal hypoglycemia, improving predinner hyperglycemia, and reducing within-day glucose variability in subjects with T1DM.

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