

6. Comi G, Sora MGN, Bianchi A, Bontempi B, Gianoglio P, Cerutti S, Micossi P, Canal N: Spectral analysis of short-term heart rate variability in diabetic patients. *J Auton Nerv Syst* 30:S45–S50, 1990

Prevalence of a Polymorphism of the Phosphatidylinositol 3-Kinase p85 α Regulatory Subunit (Codon 326^{Met}→^{Ile}) in Japanese NIDDM Patients

Phosphatidylinositol 3-kinase is a heterodimeric enzyme composed of an 85-kDa regulatory subunit (p85), containing two SH2 and one SH3 domain, and a 110-kDa catalytic subunit (p110) (1,2). This kinase has been shown to be important for insulin-stimulated glucose transport in insulin sensitive cells (3–5).

Recently, Andersen et al. (6) identified a missense mutation at nucleotide 1020 (G→A) changing Met to Ile (codon 326^{Met}→^{Ile}) in the p85 α regulatory subunit of phosphatidylinositol 3-kinase. Thus, we examined the prevalence of a polymorphism of codon 326^{Met}→^{Ile} of the p85 α subunit of PI 3-kinase, using the double mismatched polymerase chain reaction (PCR)–restriction fragment length polymorphism (RFLP) analysis (Fig. 1) to understand the contribution of this mutation to the development of NIDDM in Japanese patients. The diagnosis of NIDDM and the definition of normal glucose tolerance were established, following a 75-g oral glucose tolerance test, based on the guidelines of the World Health Organization. We examined 260 normal control subjects (mean age, 57.0 \pm 10.5 years; BMI, 22.6 \pm 2.6 kg/m²) and 200 Japanese NIDDM patients (mean age, 56.8 \pm 13.7 years; BMI, 24.9 \pm 5.2 kg/m²). In agreement with the results of Andersen et al. (6), there was no significant difference between the normal subjects and the NIDDM patients in the distribution of the wild-type, heterozygote, and homozygote genotypes (198, 57, and 5 in normal subjects vs. 150, 43, and 7 in NIDDM patients, respectively) or in the allelic frequency (12.9% in normal subjects vs. 14.3% in NIDDM patients, respectively). We also compared the clinical characteristics of

each genotype (wild-type, heterozygote, and homozygote) among the NIDDM patients and found that several parameters, including BMI (25.3 \pm 4.7, 23.9 \pm 6.8, and 26.2 \pm 3.6 kg/m²), fasting glucose (143.2 \pm 39.9, 156.7 \pm 53.8, and 157.0 \pm 111.7 mg/dl), and age at onset of diabetes (46.1 \pm 15.0, 47.1 \pm 11.4, and 47.5 \pm 10.8 years) were not statistically different among the three genotypes.

In conclusion, our results suggest that the amino acid polymorphism at codon 326^{Met}→^{Ile} in the p85 α regulatory subunit of phosphatidylinositol 3-kinase is unlikely to be a major contributor to the pathogenesis of NIDDM in Japanese patients. However, further studies are necessary to determine whether the mutations of phosphatidylinositol 3-kinase are involved in the pathogenesis of NIDDM.

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Wild Type DNA	GTAGCCAACAACGGTATGAATAACAATATGTCCTTACAAAATGCTGAAT
Sense primer	5'- GTAGCCAACAACGGTATGAATAACCATAT -3'
	↓
PCR product	GTAGCCAACAACGGTATGAATAACCATATGTCCTTACAAAATGCTGAAT
	Nde I site
Mutant DNA	GTAGCCAACAACGGTATGAATAACAATATATTCCTTACAAAATGCTGAAT
Sense primer	5'- GTAGCCAACAACGGTATGAATAACAAGAT -3'
	↓
PCR product	GTAGCCAACAACGGTATGAATAACCATATTCCTTACAAAATGCTGAAT
	EcoR V site
Antisense primer	5'- AATGGAACCTTAGCAAGCTGGTGCT -3'

Figure 1—Oligonucleotide sequences of DNA primers used for the PCR-RFLP analysis. Bases with dotted tops are the mismatched bases for the PCR-RFLP analysis. The products of the PCR fragments amplified from wild-type and mutant DNA create the restriction enzyme sites for NdeI and EcoRV, respectively. The created enzyme sites are underlined. The antisense primer is settled in the intron. The mutated base is double-underlined.

References

- Skolnik EY, Margolis B, Mohammadi M, Lowenstein E, Fischer R, Drepps A, Ullrich A, Schlessinger J: Cloning of PI 3 kinase-associated p85 utilizing a novel method for expression/cloning of target proteins for receptor tyrosine kinases. *Cell* 65:83–90, 1991
- Otsu M, Hiles I, Gout I, Fry MJ, Ruiz-Larrea F, Panayotou G, Thompson A, Dhand R, Hsuan J, Totty N, Smith AD, Morgan SJ, Courtneidge SA, Parker PJ, Waterfield MD: Characterization of two 85 kd proteins that associate with receptor tyrosine kinases, middle-T/pp60^{c-src} complexes, and PI 3-kinase. *Cell* 65:91–104, 1991
- Hara K, Yonezawa K, Sakaue H, Ando A, Kotani K, Kitamura T, Kitamura Y, Ueda H, Stephens L, Jackson TR, Hawkins PT, Dhand R, Clark AE, Holman GD, Waterfield MD, Kasuga M: 1-phosphatidylinositol 3-kinase activity is required for insulin-stimulated glucose transport but not for RAS activation in CHO cells. *Proc Natl Acad Sci USA* 91:7415–7419, 1994
- Cheatham B, Vlahos CJ, Cheatham L, Wang L, Blenis J, Kahn CR: Phosphatidylinositol 3-kinase activation is required for insulin stimulation of pp70 S6 kinase, DNA synthesis, and glucose transporter translocation. *Mol Cell Biol* 14:4902–4911, 1994
- Okada T, Kawano Y, Sakakibara T, Hazeki O, Ui M: Essential role of phosphatidylinositol 3-kinase in insulin-induced glucose transport and antilipolysis in rat adipocytes. *J Biol Chem* 269:3568–3573, 1994
- Hansen T, Andersen CB, Echwald SM, Urhammer SA, Clausen JO, Vestergaard H, Owens D, Hansen L, Pedersen O: Identification of a common amino acid polymorphism in the p85 α regulatory subunit of phosphatidylinositol 3-kinase. *Diabetes* 46:494–501, 1997